# Developing computational strategies for assembly of heterozygous DNA sequence data 

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The process of genome assembly, recently possible due to technological advancements, has presented large amounts of genetic sequence data. However, DNA must first be split into many small fragments which must be read, compared and merged to recover the original genome sequence. Specifically focusing on Corylus avellana, also known as the hazelnut, the focus of the project is to work with a large whole-genome sequence dataset, a diploid genome with high heterozygosity, to determine the original genome sequence. Multiple existing programs and software were used to develop new solutions to solve issues of highly heterozygous genomes and create a more integrated and holistic genome assembly.

## OBJECTIVES

The purpose of this project was to develop a strategy for the assembly of highly heterozygous genomes, specifically working with data of the Corylus avellana cv. Tombul. Initial genome assembly of Corylus avellana, also known as European hazelnut, produced large numbers of duplicated elements and a larger than expected genome size, implying problems due to heterozygosity. Working on the large whole-genome sequence data and using various existing tools and different software programs, the goal of the project was to solve issues of heterozygosity and develop new ways to filter and present data for a more complete genome assembly.
Through numerous data filtering and analysis procedures, by the end of the project, a data table was created to show the different areas of heterozygosity, with the information of the nucleotide, type of variation (insertion, deletion or single nucleotide polymorphism(SNP)) and starting and ending position of the heterozygous section, each matched with a specific consensus ID.

## PROJECT DETAILS

1. Genome Assembly Method In order to assemble our haplotypes, find out homozygous regions where the sequences are likely to be assembled into a continuous string, and heterozygous regions where there are multiple ways that a continuous string can be formed, we have used Platanus-allee with Nanopore reads and Illumina reads as inputs. The algorithm of Platanus-allee uses De Bruijn graphs to assemble reads into contigs using optimized kmers in this process. Kmers are small words of length $k$ observed more than once in a genomic sequence. Nodes represent kmers and edges represent k-1 overlaps between kmers. Differently than prior assemblers, Platanus automatically extends kmers to handle big and repetitive data. Once contigs are formed, they are scaffolded based on paired end libraries or mate pair libraries. In these contig assembly and scaffolding steps, complicated graph structures are simplified. Contig and scaffold construction is based on graphs without junctions; that is if a node has multiple edges.
2. Graph of heterozygous areas With the data set obtained from the Platanus-allee, the result was visualized with a graph using a logarithmic scale. X -axis represents the frequency of the kmers and y axis represents the number of unique kmers. The graph shows areas of heterozygosity in the output by comparing areas of overlap in frequency of the kmers against the number of unique kmers.

3. File Production for genome analysis and assembly The first step included using "bwa- BurrowsWheeler Alignment Tool": a software package that consists of different algorithms to map lowdivergent sequences with a large reference genome (Heng, 2010). This created a .SAM output file, which was further processed and converted to a Binary Alignment/Map (BAM) file using samtools. Samtools uses the Sequence Alignment/Map file format to sort, merge, index and retrieve reads in any region, and is able to import or export files in both SAM and BAM format(Center for Statistical Analysis, 2010). Once the sorted BAM file was created, bcftools - a tool for Binary Call Format (BCF) and VCF - was finally used to create a VCF file.

## PROJECT DETAILS

4. Graphical Assistance Throughout this process, another software application called "Tablet" was used to help give further understanding, providing a more visual representation of the sequence alignment map. It revealed the depth of the genome sequence and the areas of overlap and places of potential insertion/deletion to determine the heterozygous parts of the sequence genome (Milne et al, 2013).


FINAL RESULTS

The final table given on the right indicates the starting and ending positions of the heterozygous sites. This table was constructed using 6376 SNVs, each ranking Phred score of at least 30, so only one in a 1000 SNVs may be an artifact. An additional table was constructed using more than 2600000 SNVs scoring a Phred score more than 20. We have looked at whether the SNVs were much closer than we would expect if they were to be randomly distributed rather than being concentrated in a heterozygous region. If they were particularly concentrated in one place, the beginning and end position of the region would be noted and put in the table on the left


## CONCLUSIONS

By the end of the project, we were able to successfully identify the heterozygous regions in the genome. The dataset we have used included SNP's with a Phred score above 30: only one in every 1000 reported SNVs would be an error. However a large proportion of the SNVs given the final vcf data ranked a Phred score of about 25 , indicating that about one in 316 SNV reportings may actually be an error. We also constructed the table using the data of SNVs with a Phred score of at least 21; indicating one in 125 reportings may be an error. 2673571 SNVs was used for this table. The latter table may be more throughout and dependable
For further improvements, there are upcoming and developing softwares: such as ones promising to work with lower coverage, but unable to handle repetitive regions yet, or previously released softwares being improved such as Meraculous-2D (Goltsman 2017). Based on the current literature we think that haplotype sensitive genome assemblers are quickly developing and improving, and genome assemblies in the future will be much easier and dependable.
Our results were necessary for proceeding to next steps of genome assembly of Tombul cultivar of the hazelnut. Currently, we have yet to find out what haplotypes have got which heterozygous regions. In conclusion, using specific conditions during the filtering process, including depth and quality, the first final data table obtained was smaller than expected, which allows for further research and testing in different filtering processes and conditions to acquire a more realistic genome sequence.

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